

Mechanism of Fenfluramine in a Zebrafish Mutant Model of Dravet Syndrome?

Jo Sourbron¹, Angéla Kecskés¹, Lieven Lagae², Ilse Smolders³, Peter de Witte¹

¹Laboratory for Molecular Biodiscovery, Department of Pharmaceutical and Pharmacological Sciences, KU Leuven, Leuven, Belgium

²Department of Development and Regeneration, Section Pediatric Neurology, University Hospitals KU Leuven, Leuven, Belgium

³Center for Neurosciences, C4N, Faculty of Medicine and Pharmacy, Vrije Universiteit Brussel, Brussels, Belgium

Poster ID #2326461

(Session #3.039)



INTRODUCTION



Epilepsy and Dravet syndrome

- Epilepsy= common neurological disorder (up to 65 million people worldwide)
- 30% not responding to current anti-epileptic drugs (AEDs) (i.e. drug-resistant)
- Etiology: genetic, structural-metabolic or unknown
 - SCN1A (neural sodium channel, type 1, subunit α): most clinically relevant epilepsy gene
 - SCN1A mutation in 80% of Dravet syndrome patients
- Dravet syndrome: **drug-resistant**, rare and severe epilepsy syndrome; starting in 1st year of life, accompanied by cognitive and behavioral impairments

→ need for **new, innovative AEDs**

Zebrafish (ZF) and drug discovery

- Small vertebrate model → ZF larvae: validated for fast, cost-efficient AED screening
- Serotonin** receptors (=5-hydroxytryptamine, 5-HT-R): suggested, novel target of future AEDs²
- Experimental serotonergic drug in clinical trial for Dravet syndrome patients (**fenfluramine**):
 - Anti-epileptiform activity in *morpholino knockdown* ZF model of Dravet syndrome³
 - Severe, possible **side effects** due to stimulation of **5-HT_{2B}** receptor (valvulopathy, cardiotoxicity)
- Mechanism of action: involvement of 5-HT or other receptors?

→ from bed-side to bench and back to bed

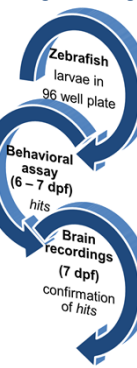
RATIONALE

How do *scn1Lab* mutant zebrafish larvae recapitulate Dravet syndrome and aid innovative anti-epileptiform drug discovery?

- Abnormal behavior and brain activity in *scn1Lab*^{-/-} mutants (-/-) vs. wildtype *scn1Lab*^{+/+} (+/+)⁴
- Drug screening of serotonergic analogs in locomotor assay leading to *hits*
- Confirmation of *hits* by measuring local field potentials and elucidating the mechanism of action

METHODS

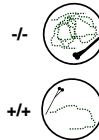
Flow Drug screening



1) Locomotor behavior and brain activity

Locomotor behavior 4 - 8 dpf

- Distance travelled in large movements for 10 minutes (min) after 30 min habituation
- High throughput (**HT**) with an automated tracking device (ZebraBox™ apparatus; Viewpoint, Lyon, France)



Brain activity 7 dpf

- Invasive forebrain open-field recordings for 10 min
- Low throughput testing (**LT**), ZF larva embedded in 2% low-melting-point agarose (Digidata 1440A digitizer; Axon instruments, USA)

2) Neurotransmission

LC-ECD 7 dpf

Determination of the amount of neurotransmitters in heads of ZF larvae by Microbore Liquid Chromatography with Electrical Chemical Detection (LC-ECD)⁵

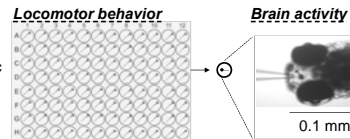
3) Drug screening

Locomotor behavior 6 - 7 dpf

- Treatment of fenfluramine and serotonergic agonists at maximum tolerated concentration (MTC)
- Combined treatment of fenfluramine and serotonergic antagonists at MTC

Brain activity 7 dpf

Confirmation of *hits* if a significant decrease in epileptiform brain activity

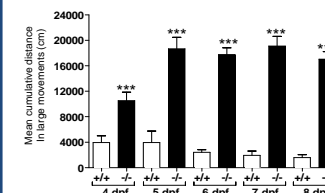


RESULTS

1) Behavior and brain activity

Locomotor behavior 4 - 8 dpf

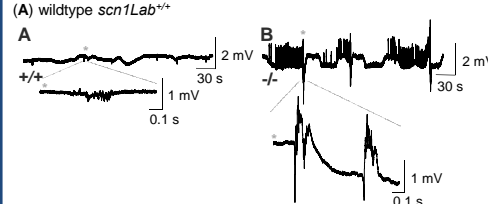
Higher activity of *scn1Lab*^{-/-} mutants, compared to wildtype *scn1Lab*^{+/+} = may reflect tonic-clonic onset of seizures in Dravet syndrome patients



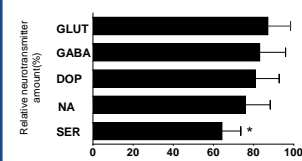
Quantification of distance in large movements (lardest) of ZF larvae. Black bars represent *scn1Lab*^{-/-} mutants and white bars represent wildtype *scn1Lab*^{+/+} (24 larvae on each day, experiment in triplicate)

Brain activity 7 dpf

Epileptiform brain activity (representative 5 min recording) = amplitude >3 times the background noise and duration >50 milliseconds (ms) in (B) *scn1Lab*^{-/-} mutants, compared to baseline in (A) wildtype *scn1Lab*^{+/+}



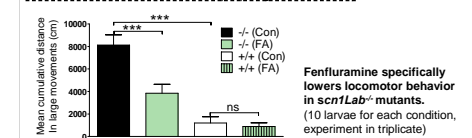
2) Neurotransmission



Decrease in serotonin in heads of *scn1Lab*^{-/-} mutants. Comparison to wildtype *scn1Lab*^{+/+} (Normalized against these age-matched controls and expressed in percentage (%)) (54 head homogenates for each condition)

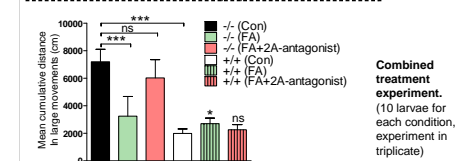
3) Drug screening

Locomotor behavior 6 - 7 dpf (single treatment)



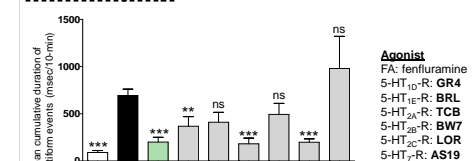
Fenfluramine specifically lowers locomotor behavior in *scn1Lab*^{-/-} mutants. (10 larvae for each condition, experiment in triplicate)

Locomotor behavior 6 - 7 dpf (combined treatment)



Counteraction of fenfluramine-induced decrease by the 5-HT_{1D}, 5-HT_{2A}, 5-HT_{2C}-antagonist

Brain activity 7 dpf



Decrease in epileptiform events. Comparison to VHC-treated *scn1Lab*^{-/-} mutants (-/-(Con)), represented by a black bar (at least 10 larvae per condition)

Fenfluramine and the 5-HT_{1D}, 5-HT_{2A} and 5-HT_{2C}-agonist decrease epileptiform brain activity in *scn1Lab*^{-/-} mutants

CONCLUSIONS

- scn1Lab*^{-/-} mutant zebrafish larvae mimic Dravet syndrome, in line with the findings of Baraban *et al.*⁴
- Reduction of **serotonin** in DS ZF larvae = highlights role of serotonin → also in animal and human studies of **drug-resistant epilepsies**⁶
- Fenfluramine, the 5-HT_{1D}, 5-HT_{2A} and 5-HT_{2C}-agonist reduced epileptiform locomotor and brain activity → **5-HT_{2B}**-receptor **not** involved!
- 5-HT_{1D}, 5-HT_{2A} and 5-HT_{2C}-antagonists counteracted the decrease by fenfluramine in locomotor assays → **5-HT_{2B}**-receptor **not** involved!

Contributes to elucidation mechanism of fenfluramine Highlights 3 serotonin receptor subtypes (5-HT_{1D}, 5-HT_{2A} and 5-HT_{2C}) as interesting targets for future innovative AEDs

1. Ceulemans B. *et al.* *Epilepsia* 53, 1131-9 (2012).
2. Löscher W, Kitgaard H, Twyman R. E & Schmidt D. *Nat. Rev. Drug Discov.* 12, 757-76 (2013).
3. Zhang Y. *et al.* *PLoS One* 10, e0125898 (2015).

4. Baraban SC, Dinday MT, Horopan GA. *Nat. Commun.* 4: 2410 (2013).
5. Sarre S. *et al.* *Methods Mol. Biol.* 72: 185-96 (1997).
6. Fonseca NC. *et al.* *Epilepsy Res.* 111: 18-25 (2015).

STATISTICS: Locomotor behavioral analyses: One-way ANOVA followed by Dunnett's multiple comparison tests; Brain activity analyses: Mann-Whitney U test; Neurotransmission: Student's t-test (passed D'Agostino & Pearson omnibus normality test). Statistically significant differences represented by (*) if $p < 0.05$, (**) if $p < 0.01$ and (***) if $p < 0.0001$